

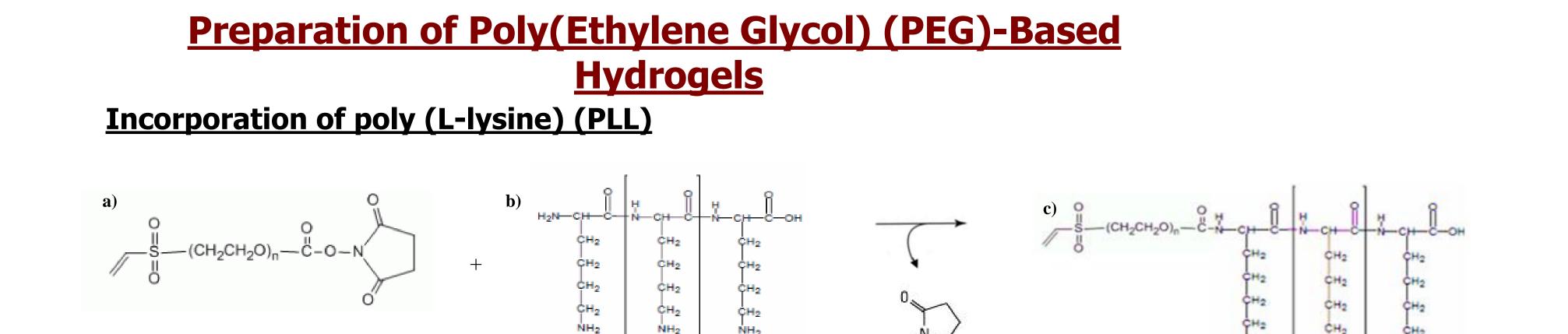
Developing a 3D Retinal Tissue Scaffold Using PEG-Based Hydrogels

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Background and Introduction

Growing interface between cell biology and bioengineering

There is a need for *an ex vivo 3D* model of the retina in order to better understand retinal disease. Standard *in vitro* cell culture methods are 2D; however, the cells in our body interact with each other in a 3D environment. Therefore, 2D cell culture methods do not provide an accurate representation of how cells interact *in vivo*. To better understand the dynamic and complex 3D extracellular environment in which our cells interact *in vivo*, biologists have been collaborating with material scientists and tissue engineers to design 3D tissue models using different types of natural and synthetic biomaterials. Our project aims to develop a method to create a 3D retinal tissue model using poly(ethylene) (PEG) based hydrogels, which are a synthetic biomaterial that can be fine-tuned to create a biomimetic scaffold of the target tissue.



Requirements for creating a tissue scaffold

- Biodegradable
- Bioactive/cell-adhesive
- Mimics mechanical properties of extracellular matrix (ECM) of target tissue
- Incorporation of signaling components that can promote a healing environment, such as celladhesion ligands, protease-sensitive domains, growth factors, and cytokines

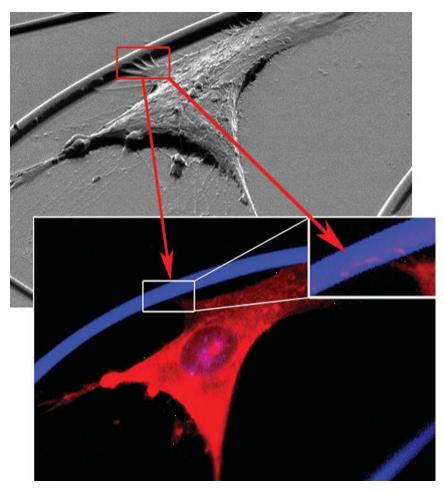


Figure 1: Image of biomimetic scaffold which favors cell attachment and growth.

Poly(ethylene glycol) (PEG) is a useful polymer to create hydrogels

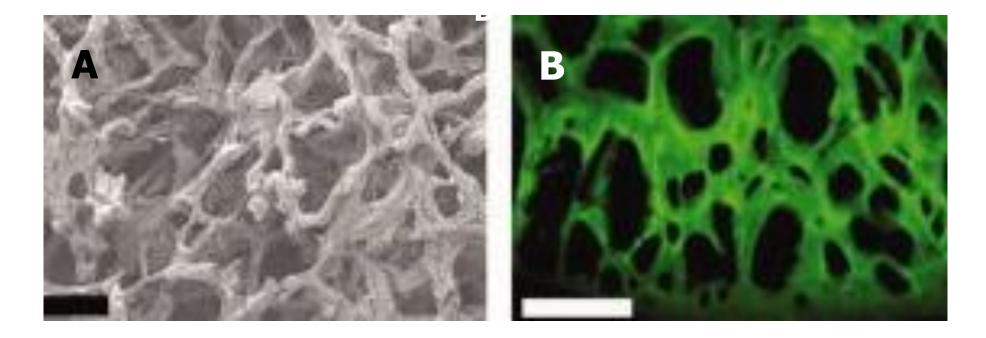


Figure 2: (a) SEM micrograph of PEG-





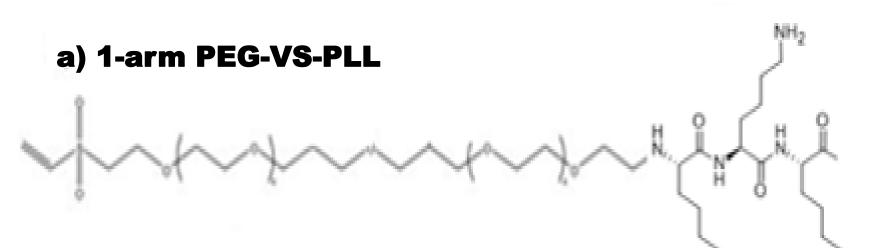
Hydrogel Casting Technique





Figure 3: Reaction scheme for incorporation of PLL. **(a)** VS-PEG-NHS polymer crosslinker mixed with **(b)** PLL in pH 10.5 buffer solution, and allowed to react for 2 hours to form **(c)** VS-PEG-PLL, a precursor for the hydrogel formation.

PEG hydrogel polymerization



b) PEG-diester-dithiol

Figure 4: Schematic of PEG hydrogel polymerization. **(a.)** 2arm PEG vinyl sulfone is dissolved in pH 8 buffer with poly (L-lysine) and **(b.)** later mixed with PEG dithiol cross linkers. The polymerization takes place in under 30 secs to form a **(c.)** 3D network.

Room Temi

PLL hydrogel. (b) 20µm cross section of PEG-PLL hydrogel reacted with FITC to demonstrate pore structure. The scale bar is equivalent for both (a) and (b).



Casting technique allows hydrogel to be produced into virtually any 3D shape

Provides the potential to produce the hydrogels into the multilayered structure of the retina in future works into different shapes to demonstrate versatility of casting technique

Hydrogels make excellent candidates for creating tissue scaffolds because they are biodegradable, can be fine-tuned to mimic the extracellular matrix (ECM) of the target tissue, have a highly porous architecture, and can be rendered bioactive.

Research Objectives

- Develop a method to synthesize a poly (ethylene) (PEG)-based hydrogel
 that is fast and does not require UV light
- Incorporate poly (L-lysine) into the hydrogel to make it cell-adhesive
- Develop a technique to produce the hydrogel into the 3D, multi-layered structure of the retina

Methods

A schematic of the hydrogel formation is presented in Figures 3 and 4. PEG is an inert substance and was made cell-adhesive by incorporating poly (L-lysine) (PLL) into the hydrogel. In order to achieve this, we mixed the PLL with VS-PEG-NHS, a polymer precursor for forming the PEG hydrogel crosslinking reaction (Figure 3). The VS-PEG-NHS and the PLL were each dissolved in phosphate buffer (PBS) at 10% w/v. The two polymers were mixed at a VS/PLL 1:1 ratio. 3M NaOH was added to obtain a final pH of 10.5, the pKa of the lysine side chain. The solutions were vortexed, and then left for 2 hours to react, and subsequently lyophilized. Then the hydrogel was formed via a Michael-type addition of PEG-diester-dithiol onto the 1-arm VS-PEG-PLL polymer (Figure 4). Each polymer precursor was dissolved in a 0.3 M triethanolamine (TEA) solution of pH 8 and 20% w/v. The ratio of VS/SH was 1:1. Immediately after mixing, the solution was vortexed for 15 secs and quickly transferred to the center of a glass slide, and casted into a specific shape (Figure 5). Gelation occurred in under 30 secs at room temperature.

Future Work

- Characterize the mechanical properties of the PEG-PLL gel
- Culture retinal progenitor cells (RPCs) on the hydrogel
 Utilize the casting technique to produce the hydrogels into the multilayered structure of the retina

Conclusion

- Synthesized VS-PEG-SH hydrogel via Michael-type addition reaction in under 30 seconds without exposure to UV light
 Rendered PEG hydrogel bioactive through incorporation of PLL
- Rendered PEG hydrogel bloactive through incorporation of PLL into the PEG-VS hydrogel precursor
- Developed casting technique to produce hydrogel into virtually any desired shape

References

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Acknowledgements

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